

Chapter 8.5

Global level: A multi-layered model to describe HIV-1 transmission

Multi-layered model to describe global HIV-1 transmission

In order to understand the HIV-1 pandemic and design preventive therapies, it is essential to further define HIV-1 transmission. From literature it is clear that there is confusion and debate on the mechanisms that are involved in the global spread of HIV-1, including the primary target cells and receptors^{7,25,52,56}. Based on previous chapters, we postulate that global sexual transmission of HIV-1 is not a static process that involves one type of target cell and receptors. We propose a multi-layered model to represent HIV-1 transmission, predict the likelihood of infection, define the first target cell and receptors involved, and to ultimately design effective methods to prevent transmission (Table 8.5; Figure 8.5).

Layer 0: Sexual practice and resistance factors

A key factor influencing sexual HIV-1 transmission in the global population is unsafe receptive intercourse. Furthermore, sex with multiple partners and traumatic sex largely increase sexual HIV-1 transmission. Resistance factors, such as CCR5 mutations⁶³, also affect HIV-1 transmission, however most of these factors are still undetermined^{63,66,92}. Awareness of safe-sex, distribution of condoms and easy access to HIV-1 test-facilities can still be improved to increase global protection against HIV-1.

1st layer: Viral load, Langerin polymorphisms, body fluid factors

High viral loads increase HIV-1 transmission. The viral load of the partner increases HIV-1 transmission by saturating Langerin function, by increasing the chance of bypassing the epithelial layers and by increasing the likelihood of infection of other target cells. Furthermore, Langerin polymorphisms decrease the interaction of this C-type lectin with carbohydrate structures¹¹⁰ and this will decrease protection by the LC barrier. Furthermore, compounds in body fluids might affect the function of the C-type lectins (Chapter 8.4). Antiretroviral therapy decreases plasma viral loads and as such decreases the susceptibility of the population to HIV-1.

Table 8.5 Multi-layered HIV-1 transmission model

Layer 0: Safe-sex/ resistance factors	Layer 1: Viral load partner	Layer 2: Exposed tissue	Layer 3: Genital “fitness”	Transmission	First target cell	Receptors involved	
NO/NO	Low	• Rectum • Endocervix	Steady-state	+	Epithelial <i>transmigration/transcytosis</i> and DC-SIGN ⁺ -DCs	HSPGs, DC-SIGN, ?CD4/CCR5/CXCR4 ? GalCer	
							++
	High	• Rectum • Endocervix	Trauma/Inflammation	++	DC-SIGN ⁺ -DCs	HSPGs, DC-SIGN, ?CD4/CCR5/CXCR4 ? GalCer	
							+++
	Low	High		Steady-state	-	-	Langerin protects
	High			Ulcerative inflammation, loss epithelium, loss LCs (hormones) or disruption epithelium (trauma)	+++	LCs	? MR/GalCer
	Low	High	• Anus • Vagina • Ectocervix • Foreskin	Non-Ulcerative Inflammation	+	LCs, DC-SIGN ⁺ DCs, subepithelial Macrophages, T cells	HSPGs, DC-SIGN, ?CD4/CCR5/CXCR4 ? MR/GalCer

2nd layer: Exposure site

Penile-anal and penile-vaginal intercourse expose different tissues to HIV-1, affecting HIV-1 transmission and the first target cells and receptors involved. During anal sex, HIV-1 encounters the rectal epithelium, and this will probably result in HIV-1 capture by DC-SIGN⁺ DCs. Vaginal and cervical tissues contain different target cells, including Langerhans cells and CD4⁺ T cells, of which the densities are highly variable and which are thought to be subject to environmental triggers. Circumcision inhibits the likelihood to acquire HIV-1²³. This implicates that the most sensitive tissue for HIV-1 transmission on the penis is the foreskin epithelium. Strikingly, the density of target cells, including LCs, is comparable in foreskin epithelium and glans epidermis, suggesting that the amount of target cells does not determine the higher susceptibility of foreskin. However, foreskin epithelium lacks the protective keratin layer of skin⁷³ and due to the mucosal structure, the epithelial/LC barrier is easier traumatized, allowing interaction of the virus with the cells in the subepithelium. Penile-vaginal and penile-anal contact may require specific microbicides to prevent HIV-1 transmission via the anal or the vaginal route.

3rd layer: Genital “fitness”

Under steady-state conditions the epithelial layer protects against HIV-1 transmission. However, under inflammatory conditions or during trauma the epithelial barrier can be disrupted, exposing subepithelial target cells to HIV-1. Furthermore, inflammation may attract target cells towards the genital epithelia and inflammatory factors might enhance HIV-1 transmission by LCs (**Section 6**) and other target cells²³. In the 1990s two major clinical trials investigated whether treatment of sexual transmitted infections (STI) reduced HIV-1 transmission. In one trial HIV-1 transmission was decreased by 38%. Unexpectedly, the other trial only marginally affected the number of HIV-1 infections^{38,42,111}. Low incidence of STIs and low-risk behaviour are thought to have attributed to the results in the second trial. Therefore, it is now thought that in population with high-risk behaviour and high STI rates, STI treatment might significantly reduce HIV-1 transmission^{62,112}.

Future directions to investigate HIV-1 using the multi-layered model

Population genetics. Population genetics have a great potential in providing answers about the involvement of different proteins in sexual HIV-1 transmission. Distinctions are needed to assess a role of different factors according to layer 1-3, comparing homosexual with heterosexual cohorts, viral loads of the infected partner and people suffering from genital inflammation compared with healthy persons.

In vivo. Primate infection with immune deficiency viruses could be a valuable mode to assess the role of the different layers by comparing the rectal, vaginal and penile route of infection, using different viral concentrations, in steady-state conditions and inflammatory conditions. In previous work, different methods were used to achieve high rates of infection using limit animal group sizes, including high viral loads and hormonal treatments. This high infectivity does not reflect the *in vivo* chance to acquire HIV-1 by sexual contact and may underestimate protective factors *in vivo*. Recently, a research group demonstrated efficient infection of macaque by SHIV using repeated low-dose exposures during several weeks¹⁰⁰. This may provide a valuable model to investigate viral transmission. Small animals are not susceptible to infection by HIV or SIV. Progress has been made towards a genetically engineered murine model that is susceptible to HIV-1 via the anal and vaginal route¹⁰. If successful, these models could provide interesting answers using knock-out for genes and cells involved in transmission^{9,50,55}.

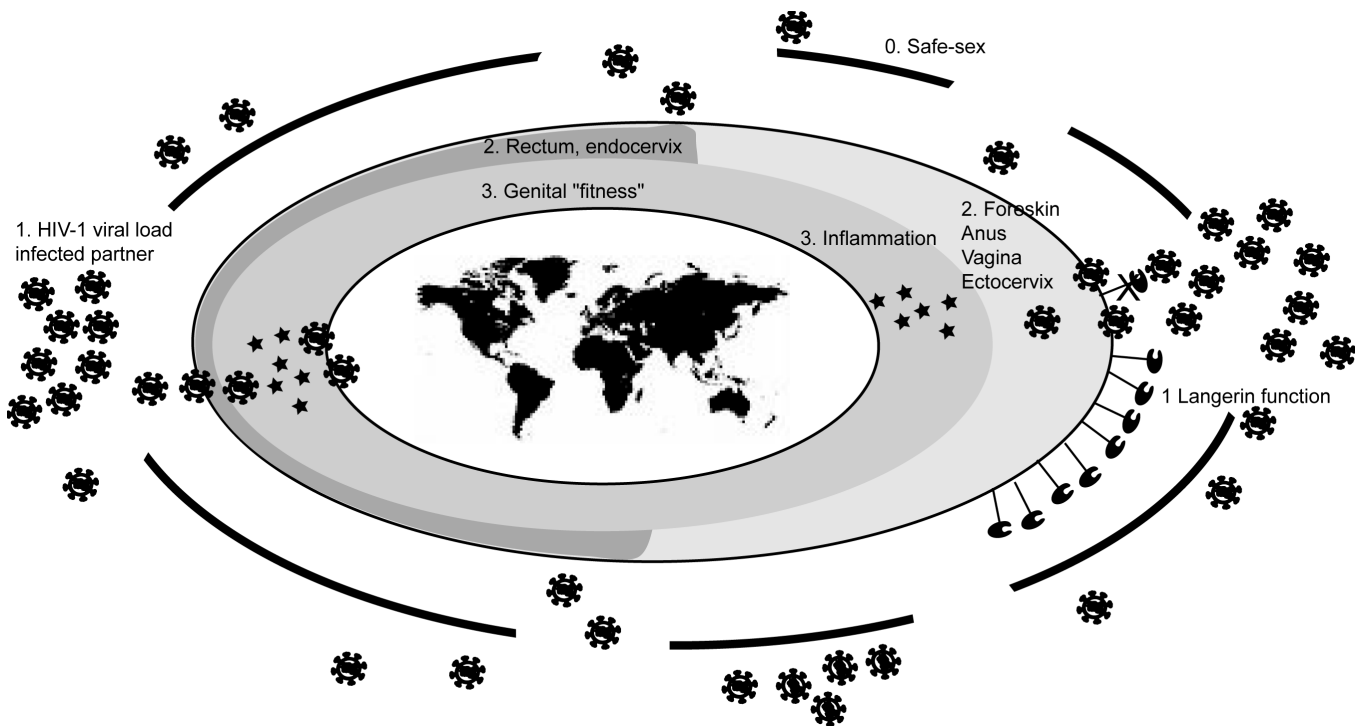


Figure 8.5 A multi-layered model to describe global HIV-1 transmission.

HIV-1 transmission is a multifactorial process. (0) Safe-sex is a major protective variable in the global population and increasing awareness on safe-sex will highly decrease HIV-1 transmission. (1) Viral load of the infected partner is strongly associated with HIV-1 transmission. High viral loads saturate the protective Langerin function and other processes that mediate transmission are enhanced. Moreover, Langerin polymorphisms or soluble factors in body fluids might affect the function of this receptor. (2) The tissue exposed to HIV-1 determines the mechanisms that are involved in HIV-1 transmission. In the rectum and endocervix the LC barrier is not present and DC-SIGN⁺ DCs that efficiently mediate transmission are closely located to the lumen. In the foreskin, vagina and ectocervix, Langerin function clears invading particles protecting the host. In some females, however, high amounts of CD4⁺ T cells are observed in the epithelia and subepithelia of the genital tissues. (3) Inflammation in all genital tissues increases HIV-1 transmission by the interaction of microbes with TLRs and the production of cytokines. Furthermore, the epithelium might be disrupted and target cells attracted towards the epithelium.

In vitro studies using primary or DC-like single cells are essential to discover new receptors and pathways. The receptors and factors that are revealed using these single cells might be essential targets for microbicides. It is important to always confirm these targets and pathways in primary cells, using different physiological concentrations of virus. More physiological concentrations of virus reveal a protective function for the C-type lectin Langerin, which is not observed using higher viral concentrations. *In vitro* studies may implement the different layers by isolation of primary cells from different target tissue, by stimulating cells with different stress-factors and by using different virus concentrations.

Design of a sensitive viral transmission model.

To ultimately draw conclusions on the first target cell and the pathways involved during transmission of HIV-1, it is essential to develop and use models that translate to the human *in vivo* situation. Transmission of the low-infectious HIV-1 is thought to be mediated by a one hit event. Unfortunately, we cannot sit down with a 'super'-microscope that visualizes this event in the human genital tract, including the identity of the cell that is targeted and the biological processes involved. This is further complicated since *trans*-infection is a process that can only be visualized using the *trans*-infected cell as a read-out. The sensitivity of our models can be increased using recombinant viruses, such as the work of Hladik *et al.* in which they infected vagina explants with HIV-1-GFP⁴³. However, this group used high concentrations of this recombinant virus and spinoculation, a method to increase infection, demonstrating that only the use of recombinant viruses is not enough to work under physiological conditions. Multiple exposure of the HIV-1-susceptible mouse²³ to physiological concentrations of HIV-1 in combination with knock-out of single HIV-1 target cell types, might reveal the contribution of the different cell types. However, the translation of this murine model to humans is not evaluated yet.

References

1989. Risk factors for male to female transmission of HIV. European Study Group. *BMJ* **298**:411-415.
1992. Comparison of female to male and male to female transmission of HIV in 563 stable couples. European Study Group on Heterosexual Transmission of HIV. *BMJ* **304**:809-813.
2003. Current Topics in Microbiology and Immunology: Dendritic Cells and Virus Infection. Springer-Verlag.
- Allan, R. S., C. M. Smith, G. T. Belz, A. L. van Lint, L. M. Wakim, W. R. Heath, and F. R. Carbone.** 2003. Epidermal viral immunity induced by CD8alpha+ dendritic cells but not by Langerhans cells. *Science* **301**:1925-1928.
- Appelmelk, B. J., I. Van Die, S. J. van Vliet, C. M. Vandenbroucke-Grauls, T. B. Geijtenbeek, and Y. van Kooyk.** 2003. Cutting edge: carbohydrate profiling identifies new pathogens that interact with dendritic cell-specific ICAM-3-grabbing nonintegrin on dendritic cells. *J. Immunol.* **170**:1635-1639.
- Atabani, S. F., A. A. Byrnes, A. Jaye, I. M. Kidd, A. F. Magnusen, H. Whittle, and C. L. Karp.** 2001. Natural measles causes prolonged suppression of interleukin-12 production. *J. Infect. Dis.* **184**:1-9.
- Balzarini, J. and L. van Damme.** 2007. Microbicide drug candidates to prevent HIV infection. *Lancet* **369**:787-797.
- Bashirova, A. A., T. B. Geijtenbeek, G. C. van Duijnhoven, S. J. van Vliet, J. B. Eilering, M. P. Martin, L. Wu, T. D. Martin, N. Viebig, P. A. Knolle, V. N. KewalRamani, K. Y. van, and M. Carrington.** 2001. A dendritic cell-specific intercellular adhesion molecule 3-grabbing nonintegrin (DC-SIGN)-related protein is highly expressed on human liver sinusoidal endothelial cells and promotes HIV-1 infection. *J. Exp. Med.* **193**:671-678.
- Bennett, C. L. and B. E. Clausen.** 2007. DC ablation in mice: promises, pitfalls, and challenges. *Trends Immunol.* **28**:525-531.
- Berges, B. K., S. R. Akkina, J. M. Folkvord, E. Connick, and R. Akkina.** 2008. Mucosal transmission of R5 and X4 tropic HIV-1 via vaginal and rectal routes in humanized Rag2(-/-)gammac(-/-) (RAG-hu) mice. *Virology*.
- Bienzle, D., K. S. Macdonald, F. M. Smill, C. Kovacs, M. Baqi, B. Courssaris, M. A. Luscher, S. L. Walmsley, and K. L. Rosenthal.** 2000. Factors contributing to the lack of human immunodeficiency virus type 1 (HIV-1) transmission in HIV-1-discordant partners. *J. Infect. Dis.* **182**:123-132.
- Bobardt, M. D., U. Chatterji, S. Selvarajah, S. B. Van der, G. David, B. Kahn, and P. A. Gallay.** 2007. Cell-free human immunodeficiency virus type 1 transcytosis through primary genital epithelial cells. *J. Virol.* **81**:395-405.
- Bobardt, M. D., A. C. Saphire, H. C. Hung, X. Yu, B. Van der Schueren, Z. Zhang, G. David, and P. A. Gallay.** 2003. Syndecan captures, protects, and transmits HIV to T lymphocytes. *Immunity* **18**:27-39.
- Bomsel, M.** 1997. Transcytosis of infectious human immunodeficiency virus across a tight human epithelial cell line barrier. *Nat Med* **3**:42-7.
- Brinkhof, M. W., J. Boni, F. Steiner, Z. Tomasik, D. Nadal, and J. Schupbach.** 2006. Evaluation of p24-based antiretroviral treatment monitoring in pediatric HIV-1 infection: prediction of the CD4+ T-cell changes between consecutive visits. *J. Acquir. Immune. Defic. Syndr.* **41**:557-562.
- Burastero, S. E., D. Gaffi, L. Lopalco, G. Tambussi, B. Borgonovo, S. C. De, C. Abecasis, P. Robbioni, A. Gasparri, A. Lazzarin, F. Celada, A. G. Siccardi, and A. Beretta.** 1996. Autoantibodies to CD4 in HIV type 1-exposed seronegative individuals. *AIDS Res. Hum. Retroviruses* **12**:273-280.
- Cambi, A., M. Koopman, and C. G. Figdor.** 2005. How C-type lectins detect pathogens. *Cell Microbiol.* **7**:481-488.
- Carreno, M. P., N. Chomont, M. D. Kazatchkine, T. Irinopoulou, C. Krief, A. S. Mohamed, L. Andreoletti, M. Matta, and L. Belec.** 2002. Binding of LFA-1 (CD11a) to intercellular adhesion molecule 3 (ICAM-3; CD50) and ICAM-2 (CD102) triggers transmigration of human immunodeficiency virus type 1-infected monocytes through mucosal epithelial cells. *J. Virol.* **76**:32-40.
- Carrillo-Farga, J., A. Castell, A. Perez, and A. Rondan.** 1990. Langerhans-like cells in amphibian epidermis. *J. Anat.* **172**:39-45.
- Chang, T. L. and M. E. Klotman.** 2004. Defensins: natural anti-HIV peptides. *AIDS Rev.* **6**:161-168.

21. **Chatwell, L., A. Holla, B. B. Kaufer, and A. Skerra.** 2007. The carbohydrate recognition domain of Langerin reveals high structural similarity with the one of DC-SIGN but an additional, calcium-independent sugar-binding site. *Mol. Immunol.*
22. **Curtis, B. M., S. Schamowski, and A. J. Watson.** 1992. Sequence and expression of a membrane-associated C-type lectin that exhibits CD4-independent binding of human immunodeficiency virus envelope glycoprotein gp120. *Proc Natl Acad Sci U S A* **89**:8356-60.
23. **de Witte, L., A. Nabatov, M. Pion, D. Fluitsma, M. A. de Jong, T. de Gruijl, V. Piguet, Y. van Kooyk, and T. B. Geijtenbeek.** 2007. Langerin is a natural barrier to HIV-1 transmission by Langerhans cells. *Nat. Med.* **13**:367-371.
24. **de Witte, L., P. A., M. D. Bobardt, A. Chatterji, U. Chatterji, J. H. Elder, G. David, S. Zolla-Pazner, M. Farzan, T. H. Lee, and P. A. Gallay.** 2005. A highly conserved arginine in gp120 governs HIV-1 binding to both syndecans and CCR5 via sulfated motifs. *J. Biol. Chem.* **280**:39493-39504.
25. **de Witte, L., A. Nabatov, and T. B. Geijtenbeek.** 2007. Distinct roles for DC-SIGN(+)-dendritic cells and Langerhans cells in HIV-1 transmission. *Trends Mol. Med.*
26. **Dong, C., A. M. Janas, J. H. Wang, W. J. Olson, and L. Wu.** 2007. Characterization of human immunodeficiency virus type 1 replication in immature and mature dendritic cells reveals dissociable cis- and trans-infection. *J. Virol.* **81**:11352-11362.
27. **Engering, A., T. B. Geijtenbeek, S. J. van Vliet, M. Wijers, L. E. van, N. Demareux, A. Lanzavecchia, J. Fransen, C. G. Figdor, V. Piguet, and Y. van Kooyk.** 2002. The dendritic cell-specific adhesion receptor DC-SIGN internalizes antigen for presentation to T cells. *J. Immunol.* **168**:2118-2126.
28. **Engering, A., S. J. van Vliet, T. B. Geijtenbeek, and Y. van Kooyk.** 2002. Subset of DC-SIGN(+) dendritic cells in human blood transmits HIV-1 to T lymphocytes. *Blood* **100**:1780-6.
29. **Fahrbach, K. M., S. M. Barry, S. Ayehunie, S. Lamore, M. Klausner, and T. J. Hope.** 2007. Activated CD34-derived Langerhans cells mediate transinfection with human immunodeficiency virus. *J. Virol.* **81**:6858-6868.
30. **Flacher, V., M. Bouschbacher, E. Verronese, C. Massacrier, V. Sisirak, O. Berthier-Vergnes, B. de Saint-Vis, C. Caux, C. Zutter-Dambuyant, S. Lebecque, and J. Valladeau.** 2006. Human Langerhans cells express a specific TLR profile and differentially respond to viruses and Gram-positive bacteria. *J. Immunol.* **177**:7959-7967.
31. **Ganz, T.** 2003. Defensins: antimicrobial peptides of innate immunity. *Nat. Rev. Immunol.* **3**:710-720.
32. **Garcia, E., M. Pion, A. Pelchen-Matthews, L. Collinson, J. F. Arrighi, G. Blot, F. Leuba, J. M. Escola, N. Demareux, M. Marsh, and V. Piguet.** 2005. HIV-1 trafficking to the dendritic cell-T-cell infectious synapse uses a pathway of tetraspanin sorting to the immunological synapse. *Traffic* **6**:488-501.
33. **Geijtenbeek, T. B., D. S. Kwon, R. Torensma, S. J. van Vliet, G. C. van Duijnhoven, J. Middel, I. L. Cornelissen, H. S. Nottet, V. N. KewalRamani, D. R. Littman, C. G. Figdor, and Y. van Kooyk.** 2000. DC-SIGN, a dendritic cell-specific HIV-1-binding protein that enhances trans-infection of T cells. *Cell* **100**:587-597.
34. **Geijtenbeek, T. B., S. J. van Vliet, E. A. Koppel, M. Sanchez-Hernandez, C. M. Vandenbroucke-Grauls, B. Appelmelk, and Y. van Kooyk.** 2003. Mycobacteria Target DC-SIGN to Suppress Dendritic Cell Function. *J Exp Med* **197**:7-17.
35. **Goh, W. C., J. Markee, R. E. Akridge, M. Meldorf, L. Musey, T. Karchmer, M. Krone, A. Collier, L. Corey, M. Emerman, and M. J. McElrath.** 1999. Protection against human immunodeficiency virus type 1 infection in persons with repeated exposure: evidence for T cell immunity in the absence of inherited CCR5 coreceptor defects. *J Infect. Dis.* **179**:548-557.
36. **Griffin DE.** 2007. Measles virus., p. 1551-1585. *In* H. P. Knipe DM (ed.), *Fields virology*. Lippincott Williams & Wilkins., Philadelphia.
37. **Gringhuis, S. I., J. den Dunnen, M. Litjens, B. van het Hof, Y. van Kooyk, and T. B. H. Geijtenbeek.** 2007. C-type lectin DC-SIGN modulates toll-like receptor signaling via Raf-1 kinase-dependent acetylation of transcription factor NF-kappa B. *Immunity* **26**:605-616.
38. **Grosskurth, H., F. Mosha, J. Todd, E. Mwijarubi, A. Klokke, K. Senkoro, P. Mayaud, J. Changalucha, A. Nicoll, G. ka-Gina, and .** 1995. Impact of improved treatment of sexually transmitted diseases on HIV infection in rural Tanzania: randomised controlled trial. *Lancet* **346**:530-536.
39. **Gupta, P., K. B. Collins, D. Ratner, S. Watkins, G. J. Naus, D. V. Landers, and B. K. Patterson.** 2002. Memory CD4(+) T cells are the earliest detectable human immunodeficiency virus type 1 (HIV-1)-infected cells in the female genital mucosal tissue during HIV-1 transmission in an organ culture system. *J. Virol.* **76**:9868-9876.
40. **Gurney, K. B., J. Elliott, H. Nassanian, C. Song, E. Soilleux, I. McGowan, P. A. Anton, and B. Lee.** 2005. Binding and transfer of human immunodeficiency virus by DC-SIGN+ cells in human rectal mucosa. *J. Virol.* **79**:5762-5773.
41. **Ha, E., M. J. Kim, B. K. Choi, J. J. Rho, D. J. Oh, T. H. Rho, K. H. Kim, H. J. Lee, D. H. Shin, S. V. Yim, H. H. Baik, J. H. Chung, and J. W. Kim.** 2006. Positive association of obesity with single nucleotide polymorphisms of syndecan 3 in the Korean population. *J Clin. Endocrinol. Metab* **91**:5095-5099.
42. **Hayes, R., F. Mosha, A. Nicoll, H. Grosskurth, J. Newell, J. Todd, J. Killewo, J. Rugemalila, and D. Mabey.** 1995. A community trial of the impact of improved sexually transmitted disease treatment on the HIV epidemic in rural Tanzania: 1. Design. *AIDS* **9**:919-926.
43. **Hladik, F., P. Sakchalathorn, L. Ballweber, G. Lentz, M. Fialkow, D. Eschenbach, and M. J. McElrath.** 2007. Initial events in establishing vaginal entry and infection by human immunodeficiency virus type-1. *Immunity* **26**:257-270.
44. **Hovius, J. W. R., M. A. W. P. de Jong, J. den Dunnen, M. Litjens, T. van der Poll, S. I. Gringhuis, and T. B. H. Geijtenbeek.** 2008. Salp15 Binding to DC-SIGN Inhibits Cytokine Expression by Impairing both Nucleosome Remodeling and mRNA Stabilization. *Plos Pathogen* **4**:1-13.
45. **Hu, J., M. B. Gardner, and C. J. Miller.** 2000. Simian immunodeficiency virus rapidly penetrates the cervicovaginal mucosa after intravaginal inoculation and infects intraepithelial dendritic cells. *J Virol* **74**:6087-95.
46. **Hu, Q., I. Frank, V. Williams, J. J. Santos, P. Watts, G. E. Griffin, J. P. Moore, M. Pope, and R. J. Shattock.** 2004. Blockade of Attachment and Fusion Receptors Inhibits HIV-1 Infection of Human Cervical Tissue. *J. Exp. Med.* **199**:1065-1075.

47. Huang, Y., W. A. Paxton, S. M. Wolinsky, A. U. Neumann, L. Zhang, T. He, S. Kang, D. Ceradini, Z. Jin, K. Yazdanbakhsh, K. Kunstman, D. Erickson, E. Dragon, N. R. Landau, J. Phair, D. D. Ho, and R. A. Koup. 1996. The role of a mutant CCR5 allele in HIV-1 transmission and disease progression. *Nat. Med.* **2**:1240-1243.
48. Jameson, B., F. Baribaud, S. Pohlmann, D. Ghavimi, F. Mortari, R. W. Doms, and A. Iwasaki. 2002. Expression of DC-SIGN by dendritic cells of intestinal and genital mucosae in humans and rhesus macaques. *J. Virol.* **76**:1866-1875.
49. Jendrysik, M. A., M. Ghassemi, P. J. Graham, L. A. Boksa, P. R. Williamson, and R. M. Novak. 2005. Human cervicovaginal lavage fluid contains an inhibitor of HIV binding to dendritic cell-specific intercellular adhesion molecule 3-grabbing nonintegrin. *J. Infect. Dis.* **192**:630-639.
50. Jung, S., D. Unutmaz, P. Wong, G. Sano, S. K. De los, T. Sparwasser, S. Wu, S. Vuthoori, K. Ko, F. Zavala, E. G. Pamer, D. R. Littman, and R. A. Lang. 2002. In vivo depletion of CD11c(+) dendritic cells abrogates priming of CD8(+) T cells by exogenous cell-associated antigens. *Immunity.* **17**:211-220.
51. Kawamura, T., S. S. Cohen, D. L. Borris, E. A. Aquilino, S. Glushakova, L. B. Margolis, J. M. Orenstein, R. E. Offord, A. R. Neurath, and A. Blauvelt. 2000. Candidate microbicides block HIV-1 infection of human immature Langerhans cells within epithelial tissue explants. *J. Exp. Med.* **192**:1491-1500.
52. Kawamura, T., S. E. Kurtz, A. Blauvelt, and S. Shimada. 2005. The role of Langerhans cells in the sexual transmission of HIV. *J. Dermatol. Sci.* **40**:147-155.
53. Kawamura, T., M. Qualbani, E. K. Thomas, J. M. Orenstein, and A. Blauvelt. 2001. Low levels of productive HIV infection in Langerhans cell-like dendritic cells differentiated in the presence of TGF-beta1 and increased viral replication with CD40 ligand-induced maturation. *Eur. J. Immunol.* **31**:360-368.
54. Kazmi, S. H., J. R. Naglik, S. P. Sweet, R. W. Evans, S. O'Shea, J. E. Banatvala, and S. J. Challacombe. 2006. Comparison of human immunodeficiency virus type 1-specific inhibitory activities in saliva and other human mucosal fluids. *Clin. Vaccine Immunol.* **13**:1111-1118.
55. Kissenpfennig, A., S. it-Yahia, V. Clair-Moninot, H. Stossel, E. Badell, Y. Bordat, J. L. Pooley, T. Lang, E. Prina, I. Coste, O. Gresser, T. Renno, N. Winter, G. Milon, K. Shortman, N. Romani, S. Lebecque, B. Malissen, S. Saeland, and P. Douillard. 2005. Disruption of the langerin/CD207 gene abolishes Birbeck granules without a marked loss of Langerhans cell function. *Mol. Cell Biol.* **25**:88-99.
56. Klasse, P. J., R. J. Shattock, and J. P. Moore. 2006. Which topical microbicides for blocking HIV-1 transmission will work in the real world? *PLoS. Med.* **3**:e351.
57. Klimstra, W. B., E. M. Nangle, M. S. Smith, A. D. Yurochko, and K. D. Ryman. 2003. DC-SIGN and L-SIGN can act as attachment receptors for alphaviruses and distinguish between mosquito cell- and mammalian cell-derived viruses. *J. Virol.* **77**:12022-12032.
58. Koizumi, Y., S. Kageyama, Y. Fujiyama, M. Miyashita, R. Lwembe, K. Ogino, T. Shioda, and H. Ichimura. 2007. RANTES -28G delays and DC-S. *AIDS Res. Hum. Retroviruses* **23**:713-719.
59. Komine, M., M. Karakawa, T. Takekoshi, N. Sakurai, Y. Minatani, H. Mitsui, Y. Tada, H. Saeki, A. Asahina, and K. Tamaki. 2007. Early inflammatory changes in the "perilesional skin" of psoriatic plaques: is there interaction between dendritic cells and keratinocytes? *J. Invest Dermatol.* **127**:1915-1922.
60. Koppel, E. A., I. S. Ludwig, B. J. Appelmeik, Y. van Kooyk, and T. B. Geijtenbeek. 2005. Carbohydrate specificities of the murine DC-SIGN homologue mSIGNR1. *Immunobiology* **210**:195-201.
61. Koppel, E. A., K. P. van Gisbergen, T. B. Geijtenbeek, and Y. van Kooyk. 2005. Distinct functions of DC-SIGN and its homologues L-SIGN (DC-SIGNR) and mSIGNR1 in pathogen recognition and immune regulation. *Cell Microbiol.* **7**:157-165.
62. Korenromp, E. L., R. G. White, K. K. Orroth, R. Bakker, A. Kamali, D. Serwadda, R. H. Gray, H. Grosskurth, J. D. Habbema, and R. J. Hayes. 2005. Determinants of the impact of sexually transmitted infection treatment on prevention of HIV infection: a synthesis of evidence from the Mwanza, Rakai, and Masaka intervention trials. *J. Infect. Dis.* **191 Suppl 1**:S168-S178.
63. Kulkarni, P. S., S. T. Butera, and A. C. Duerr. 2003. Resistance to HIV-1 infection: lessons learned from studies of highly exposed persistently seronegative (HEPS) individuals. *AIDS Rev.* **5**:87-103.
64. Langlade-Demoyen, P., N. Ngo-Giang-Huong, F. Ferchal, and E. Oksenhendler. 1994. Human immunodeficiency virus (HIV) nef-specific cytotoxic T lymphocytes in noninfected heterosexual contact of HIV-infected patients. *J. Clin. Invest* **93**:1293-1297.
65. Lazzarin, A., A. Saracco, M. Musicco, and A. Nicolosi. 1991. Man-to-woman sexual transmission of the human immunodeficiency virus. Risk factors related to sexual behavior, man's infectiousness, and woman's susceptibility. Italian Study Group on HIV Heterosexual Transmission. *Arch Intern. Med.* **151**:2411-2416.
66. Lederman, M. M., R. E. Offord, and O. Hartley. 2006. Microbicides and other topical strategies to prevent vaginal transmission of HIV. *Nat. Rev. Immunol.* **6**:371-382.
67. Liu, R., W. A. Paxton, S. Choe, D. Ceradini, S. R. Martin, R. Horuk, M. E. MacDonald, H. Stuhlmann, R. A. Koup, and N. R. Landau. 1996. Homozygous defect in HIV-1 coreceptor accounts for resistance of some multiply-exposed individuals to HIV-1 infection. *Cell* **86**:367-377.
68. Lopalco, L., C. Barassi, C. Pastori, R. Longhi, S. E. Burastero, G. Tambussi, F. Mazzotta, A. Lazzarin, M. Clerici, and A. G. Siccardi. 2000. CCR5-reactive antibodies in seronegative partners of HIV-seropositive individuals down-modulate surface CCR5 in vivo and neutralize the infectivity of R5 strains of HIV-1 In vitro. *J. Immunol.* **164**:3426-3433.
69. Lovy, J., G. M. Wright, and D. J. Speare. 2006. Morphological presentation of a dendritic-like cell within the gills of chinook salmon infected with Loma salmonae. *Dev. Comp Immunol.* **30**:259-263.
70. Ludwig, I. S., A. N. Lekkerkerker, E. Depla, F. Bosman, R. J. Musters, S. Depraetere, Y. van Kooyk, and T. B. Geijtenbeek. 2004. Hepatitis C virus targets DC-SIGN and L-SIGN to escape lysosomal degradation. *J. Virol.* **78**:8322-8332.
71. Martin, M. P., M. M. Lederman, H. B. Hutcheson, J. J. Goedert, G. W. Nelson, K. Y. van, R. Detels, S. Buchbinder, K. Hoots, D. Vlahov, S. J. O'Brien, and M. Carrington. 2004. Association of DC-SIGN promoter polymorphism with increased risk for parenteral, but not mucosal, acquisition of human immunodeficiency virus type 1 infection. *J. Virol.* **78**:14053-14056.

72. **Mc, D. R., U. Ziyilan, D. Spehner, H. Bausinger, D. Lipsker, M. Mommaas, J. P. Cazenave, G. Raposo, B. Goud, S. H. De La, J. Salamero, and D. Hanau.** 2002. Birbeck granules are subdomains of endosomal recycling compartment in human epidermal Langerhans cells, which form where Langerin accumulates. *Mol. Biol. Cell* **13**:317-335.
73. **McCoombe, S. G. and R. V. Short.** 2006. Potential HIV-1 target cells in the human penis. *AIDS* **20**:1491-1495.
74. **McDermott, R., H. Bausinger, D. Fricker, D. Spehner, F. Proamer, D. Lipsker, J. P. Cazenave, B. Goud, S. H. De La, J. Salamero, and D. Hanau.** 2004. Reproduction of Langerin/CD207 traffic and Birbeck granule formation in a human cell line model. *J. Invest Dermatol.* **123**:72-77.
75. **McDonald, D., L. Wu, S. M. Bohks, V. N. KewalRamani, D. Unutmaz, and T. J. Hope.** 2003. Recruitment of HIV and its receptors to dendritic cell-T cell junctions. *Science* **300**:1295-7.
76. **Meng, G., X. Wei, X. Wu, M. T. Sellers, J. M. Decker, Z. Moldoveanu, J. M. Orenstein, M. F. Graham, J. C. Kappes, J. Mestecky, G. M. Shaw, and P. D. Smith.** 2002. Primary intestinal epithelial cells selectively transfer R5 HIV-1 to CCR5+ cells. *Nat. Med.* **8**:150-156.
77. **Merad, M., N. Romani, and G. Randolph.** 2008. Langerhans cells at the interface of medicine, science, and industry. *J. Invest Dermatol.* **128**:251-255.
78. **Moris, A., C. Nobile, F. Buseyne, F. Porrot, J. P. Abastado, and O. Schwartz.** 2003. DC-SIGN promotes exogenous MHC-I restricted HIV-1 antigen presentation. *Blood.*
79. **Moris, A., A. Pajot, F. Blanchet, F. Guivel-Benhassine, M. Salcedo, and O. Schwartz.** 2006. Dendritic cells and HIV-specific CD4+ T cells: HIV antigen presentation, T cell activation, viral transfer. *Blood.*
80. **Moss, W. J., J. J. Ryon, M. Monze, and D. E. Griffin.** 2002. Differential regulation of interleukin (IL)-4, IL-5, and IL-10 during measles in Zambian children. *J Infect. Dis.* **186**:879-887.
81. **Naarding, M. A., A. M. Dirac, I. S. Ludwig, D. Speijer, S. Lindquist, E. L. Vestman, M. J. Stax, T. B. Geijtenbeek, G. Pollakis, O. Hermell, and W. A. Paxton.** 2006. Bile salt-stimulated lipase from human milk binds DC-SIGN and inhibits human immunodeficiency virus type 1 transfer to CD4+ T cells. *Antimicrob. Agents Chemother.* **50**:3367-3374.
82. **Naarding, M. A., I. S. Ludwig, F. Groot, B. Berkhout, T. B. Geijtenbeek, G. Pollakis, and W. A. Paxton.** 2005. Lewis X component in human milk binds DC-SIGN and inhibits HIV-1 transfer to CD4+ T lymphocytes. *J. Clin. Invest* **115**:3256-3264.
83. **Padian, N., L. Marquis, D. P. Francis, R. E. Anderson, G. W. Rutherford, P. M. O'Malley, and W. Winkelstein, Jr.** 1987. Male-to-female transmission of human immunodeficiency virus. *JAMA* **258**:788-790.
84. **Patton, J. C., G. G. Sherman, A. H. Coovadia, W. S. Stevens, and T. M. Meyers.** 2006. Ultrasensitive human immunodeficiency virus type 1 p24 antigen assay modified for use on dried whole-blood spots as a reliable, affordable test for infant diagnosis. *Clin. Vaccine Immunol.* **13**:152-155.
85. **Raboud, J. M., J. S. Montaner, B. Conway, L. Haley, C. Sherlock, M. V. O'Shaughnessy, and M. T. Schechter.** 1996. Variation in plasma RNA levels, CD4 cell counts, and p24 antigen levels in clinically stable men with human immunodeficiency virus infection. *J Infect. Dis.* **174**:191-194.
86. **Reece, J. C., A. J. Handley, E. J. Anstee, W. A. Morrison, S. M. Crowe, and P. U. Cameron.** 1998. HIV-1 selection by epidermal dendritic cells during transmission across human skin. *J Exp Med* **187**:1623-31.
87. **Reizes, O., D. J. Clegg, A. D. Strader, and S. C. Benoit.** 2006. A role for syndecan-3 in the melanocortin regulation of energy balance. *Peptides* **27**:274-280.
88. **Rescigno, M., M. Urbano, B. Valzasina, M. Francolini, G. Rotta, R. Bonasio, F. Granucci, J. P. Kraehenbuhl, and P. Ricciardi-Castagnoli.** 2001. Dendritic cells express tight junction proteins and penetrate gut epithelial monolayers to sample bacteria. *Nat. Immunol.* **2**:361-367.
89. **Ritter, U., A. Meissner, C. Scheidig, and H. Komer.** 2004. CD8 alpha- and Langerin-negative dendritic cells, but not Langerhans cells, act as principal antigen-presenting cells in leishmaniasis. *Eur. J. Immunol.* **34**:1542-1550.
90. **Royce, R. A., A. Sena, W. Cates, Jr., and M. S. Cohen.** 1997. Sexual transmission of HIV. *N. Engl. J. Med.* **336**:1072-1078.
91. **Sakuntabhai, A., C. Turbpaiboon, I. Casademont, A. Chuansumrit, T. Lowhnoo, A. Kajaste-Rudnitski, S. M. Kalayanarooj, K. Tangnaratchakit, N. Tangthawornchaikul, S. Vasanawathana, W. Chaiyaratana, P. T. Yenchitsomanus, P. Suriyaphol, P. Avirutnan, K. Chokephaibulkit, F. Matsuda, S. Yoksan, Y. Jacob, G. M. Lathrop, P. Malasit, P. Despres, and C. Julier.** 2005. A variant in the CD209 promoter is associated with severity of dengue disease. *Nat. Genet.* **37**:507-513.
92. **Salkowitz, J. R., S. F. Purvis, H. Meyerson, P. Zimmerman, T. R. O'Brien, L. Aledort, M. E. Eyster, M. Hilgartner, C. Kessler, B. A. Konkle, G. C. White, J. J. Goedert, and M. M. Lederman.** 2001. Characterization of high-risk HIV-1 seronegative hemophiliacs. *Clin. Immunol.* **98**:200-211.
93. **Sanders, R. W., E. C. de Jong, C. E. Baldwin, J. H. Schuitemaker, M. L. Kapsenberg, and B. Berkhout.** 2002. Differential transmission of human immunodeficiency virus type 1 by distinct subsets of effector dendritic cells. *J Virol* **76**:7812-21.
94. **Saphire, A. C., M. D. Bobardt, Z. Zhang, G. David, and P. A. Gallay.** 2001. Syndecans serve as attachment receptors for human immunodeficiency virus type 1 on macrophages. *J Virol* **75**:9187-200.
95. **Schubach, J., Z. Tomasik, M. Knuchel, M. Opravil, H. F. Gunthard, D. Nadal, and J. Boni.** 2006. Optimized virus disruption improves detection of HIV-1 p24 in particles and uncovers a p24 reactivity in patients with undetectable HIV-1 RNA under long-term HAART. *J Med. Virol.* **78**:1003-1010.
96. **Spira, A. I., P. A. Marx, B. K. Patterson, J. Mahoney, R. A. Koup, S. M. Wolinsky, and D. D. Ho.** 1996. Cellular targets of infection and route of viral dissemination after an intravaginal inoculation of simian immunodeficiency virus into rhesus macaques. *J Exp Med* **183**:215-25.
97. **Stambach, N. S. and M. E. Taylor.** 2003. Characterization of carbohydrate recognition by langerin, a C-type lectin of Langerhans cells. *Glycobiology* **13**:401-10.
98. **Takahara, K., Y. Yashima, Y. Omatsu, H. Yoshida, Y. Kimura, Y. S. Kang, R. M. Steinman, C. G. Park, and K. Inaba.** 2004. Functional comparison of the mouse DC-SIGN, SIGNR1, SIGNR3 and Langerin, C-type lectins. *Int. Immunol.* **16**:819-829.

99. **Trombetta, E. S. and I. Mellman.** 2005. Cell biology of antigen processing in vitro and in vivo. *Annu. Rev. Immunol.* **23**:975-1028.
100. **Tsai, L., N. Trunova, A. Gettie, H. Mohri, R. Bohm, M. Saifuddin, and C. Cheng-Mayer.** 2007. Efficient repeated low-dose intravaginal infection with X4 and R5 SHIVs in rhesus macaque: implications for HIV-1 transmission in humans. *Virology* **362**:207-216.
101. **Turville, S. G., P. U. Cameron, A. Handley, G. Lin, S. Pohlmann, R. W. Doms, and A. L. Cunningham.** 2002. Diversity of receptors binding HIV on dendritic cell subsets. *Nat Immunol* **3**:975-83.
102. **Uzan-Gafsou, S., H. Bausinger, F. Proamer, S. Monier, D. Lipsker, J. P. Cazenave, B. Goud, S. H. De La, D. Hanau, and J. Salamero.** 2007. Rab11A controls the biogenesis of Birbeck granules by regulating Langerin recycling and stability. *Mol. Biol. Cell* **18**:3169-3179.
103. **van der Aar, A. M., R. M. Sylva-Steenland, J. D. Bos, M. L. Kapsenberg, E. C. de Jong, and M. B. Teunissen.** 2007. Loss of TLR2, TLR4, and TLR5 on Langerhans cells abolishes bacterial recognition. *J. Immunol.* **178**:1986-1990.
104. **van Liempt E., C. M. Bank, P. Mehta, J. J. Garcia-Vallejo, Z. S. Kwar, R. Geyer, R. A. Alvarez, R. D. Cummings, Y. van Kooyk, and I. Van Die.** 2006. Specificity of DC-SIGN for mannose- and fucose-containing glycans. *FEBS Lett.* **580**:6123-6131.
105. **Veazey, R. S., P. J. Klasse, S. M. Schader, Q. Hu, T. J. Ketas, M. Lu, P. A. Marx, J. Dufour, R. J. Colonno, R. J. Shattock, M. S. Springer, and J. P. Moore.** 2005. Protection of macaques from vaginal SHIV challenge by vaginally delivered inhibitors of virus-cell fusion. *Nature* **438**:99-102.
106. **Veerman, E. C., P. A. van den Keybus, A. Vissink, and A. V. Nieuw Amerongen.** 1996. Human glandular salivas: their separate collection and analysis. *Eur. J Oral Sci* **104**:346-352.
107. **Verdijk, P., R. Dijkman, E. I. Plasmeijer, A. A. Mulder, W. H. Zoutman, M. A. Mieke, and C. P. Tensen.** 2005. A lack of Birbeck granules in Langerhans cells is associated with a naturally occurring point mutation in the human Langerin gene. *J. Invest Dermatol.* **124**:714-717.
108. **Vigerust, D. J. and V. L. Shepherd.** 2007. Virus glycosylation: role in virulence and immune interactions. *Trends Microbiol.* **15**:211-218.
109. **Wang, J. H., A. M. Janas, W. J. Olson, and L. Wu.** 2007. Functionally distinct transmission of human immunodeficiency virus type 1 mediated by immature and mature dendritic cells. *J. Virol.* **81**:8933-8943.
110. **Ward, E. M., N. S. Stambach, K. Drickamer, and M. E. Taylor.** 2006. Polymorphisms in human langerin affect stability and sugar-binding activity. *J. Biol. Chem.*
111. **Wawer, M. J., N. K. Sewankambo, D. Serwadda, T. C. Quinn, L. A. Paxton, N. Kiwanuka, F. Wabwire-Mangen, C. Li, T. Lutalo, F. Nalugoda, C. A. Gaydos, L. H. Moulton, M. O. Meehan, S. Ahmed, and R. H. Gray.** 1999. Control of sexually transmitted diseases for AIDS prevention in Uganda: a randomised community trial. Rakai Project Study Group. *Lancet* **353**:525-535.
112. **White, R. G., K. K. Orroth, E. L. Korenromp, R. Bakker, M. Wambura, N. K. Sewankambo, R. H. Gray, A. Kamali, J. A. Whitworth, H. Grosskurth, J. D. Habbema, and R. J. Hayes.** 2004. Can population differences explain the contrasting results of the Mwanza, Rakai, and Masaka HIV/sexually transmitted disease intervention trials?: A modeling study. *J Acquir. Immune. Defic. Syndr.* **37**:1500-1513.
113. World Health Organisation. measles fact sheet, No 286. 2007.
Ref Type: Report
114. **Zhang, L., T. He, A. Talal, G. Wang, S. S. Frankel, and D. D. Ho.** 1998. In vivo distribution of the human immunodeficiency virus/simian immunodeficiency virus coreceptors: CXCR4, CCR3, and CCR5. *J. Virol.* **72**:5035-5045.
115. **Zhang, Z., T. Schuler, M. Zupancic, S. Wietgreffe, K. A. Staskus, K. A. Reimann, T. A. Reinhart, M. Rogan, W. Cavert, C. J. Miller, R. S. Veazey, D. Notermans, S. Little, S. A. Danner, D. D. Richman, D. Havlir, J. Wong, H. L. Jordan, T. W. Schacker, P. Racz, K. Tenner-Racz, N. L. Letvin, S. Wolinsky, and A. T. Haase.** 1999. Sexual transmission and propagation of SIV and HIV in resting and activated CD4+ T cells. *Science* **286**:1353-7.
116. **Zhao, X., E. Deak, K. Soderberg, M. Linehan, D. Spezzano, J. Zhu, D. M. Knipe, and A. Iwasaki.** 2003. Vaginal submucosal dendritic cells, but not Langerhans cells, induce protective Th1 responses to herpes simplex virus-2. *J. Exp. Med.* **197**:153-162.